NEW ENGLAND BioLabs Inc.
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New England Biolabs
Certificate of Analysis

Product Name: *Ph.D.™-C7C Phage Display Peptide Library Kit*
Catalog Number: *E8120S*
Packaging Lot Number: *10121425*
Expiration Date: *07/2023*
Storage Temperature: *-20°C*
Specification Version: *PS-E8120S v2.0*

Ph.D.™-C7C Phage Display Peptide Library Kit Component List			
NEB Part Number	**Component Description**	**Lot Number**	**Individual QC Result**
S1259AVIAL	-96 gIII Sequencing Primer (20-mer)	10122169	**Pass**
S1258AVIAL	-28 gIII Sequencing Primer (22-mer)	10122170	**Pass**
N7024AVIAL	Biotin	10111664	**Pass**
N7023AVIAL	Streptavidin, lyophilized	10115978	**Pass**
E8121AVIAL	Ph.D.™-C7C Phage Display Peptide Library	10109413	**Pass**
E4104SVIAL	E.coli K12 ER2738	10055843	**Pass**

Assay Name/Specification	Lot # 10121425
Functional Testing (Panning) A 100-fold representation of the Ph.D.™-C7C Phage Display Peptide Library containing approximately 1011 pfu is diluted in 200 µl TBS and panned against 300 ng of anti-FLAG® monoclonal antibody. The bound phage is affinity captured using magnetic beads and eluted with 1 ml of 0.2M Glycine-HCl, pH 2.2. After three rounds of selection, ≥75% of sequences contain a motif related to the known epitope for the antibody.	**Pass**
Sequence Verification (DNA) The Ph.D.™-C7C Phage Display Peptide Library was sequenced using 5'-CCCATGTACCGTAACACTGAGTTTC-3' as a primer to confirm the correct form of the cloned insert on the displayed peptide, ACX7C-GGG.	**Pass**
Absolute Phage Titer Infection of a mid-log culture of E. coli ER2738 with Ph.D.™-C7C Phage Display Peptide Library followed by plating, yields ≥ 1 x 101^3 pfu/ml.	**Pass**
Phage Contamination (Environmental) A 1:100 dilution of an overnight culture of E. coli ER2738 was made in 20 ml LB, to which 10^3 pfu of Ph.D.™-C7C Phage Display Peptide Library was added. The flask was	**Pass**

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incubated at 37°C on a rotating shaker for 5 hours. A 1 ml volume of culture was removed and centrifuged. A volume of culture supernatant equivalent to the initial PFU input was added to a second, 20 ml culture like the first. The final culture supernatant was plated on three LB/IPTG/Xgal plates and then titered. Fewer than 20% clear or white plaques were observed in a minimum of 100 total plaques counted on each plate.	

This product has been tested and shown to be in compliance with all specifications.

One or more products referenced in this document may be covered by a 3rd-party trademark. Please visit www.neb.com/trademarks for additional information.

Alicia Bielik
Production Scientist
29 Sep 2021

Josh Hersey
Packaging Quality Control Inspector
29 Sep 2021